

Biocover Performance of Landfill Methane Oxidation: Experimental Results

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Abstract:

An experimental passive methane oxidation biocover (PMOB) was constructed within the existing final cover of the St-Nicéphore landfill. Its substrate consisted of a 0.80-m thick mixture of sand and compost. The goal of this experiment was to evaluate the performance of the PMOB in reducing CH₄ emissions when submitted to an increasing methane load. The CH₄ load applied started with 0.3 g CH₄ m⁻² h⁻¹. When the site had to be closed for the winter, the CH₄ input was 27 g CH₄ m⁻² h⁻¹. Throughout the study, practically all the CH₄ input was oxidized, absolute removal rates were linearly correlated to methane loading, and the oxidation zone was established between 0.6-0.8 m. These results seem to indicate that the upper limit potential of this PMOB to oxidize CH₄ was not reached during the study period. Surface CH₄ concentration scans showed no signs of leaks. The substrate offered excellent conditions for the growth of methanotrophs, whose count averaged 3.91 x 10⁸ CFU g dw⁻¹ soil.

CE Database subject headings: Methane; Oxidation; Landfill; Emissions.

Introduction

Methane is a radiatively active gas whose concentration in the atmosphere has increased in the last several decades due principally to the great increase in anthropogenic emissions (IPCC 2001); it is now estimated that as much as 19% of the CH₄ anthropogenic emissions to the atmosphere can be attributed to landfills (IPCC 2001). A number of design advances, such as gas collection systems, have reduced the environmental impacts of new landfills. However, in small and old landfills, gas collection is not cost effective, and as a result all the biogas produced is allowed to escape into the atmosphere, constituting what is called fugitive emissions. In addition, residual emissions are expected to be released from landfills after gas collection systems are turned off. However low they might be, such residual emissions may occur for several decades.

Methane emissions from landfills, particularly fugitive and residual emissions, can be reduced through microbial methane oxidation in landfill cover soils (Ait-Benichou et al. 2009; Hilger et al. 2000) - or biocovers. This relies principally on the activity of methane oxidizing bacteria, or methanotrophs, which are able to use molecular oxygen to oxidize CH₄ to CO₂ and cell carbon (Hanson and Hanson 1996). Biocovers are cited in the IPCC Working Group III assessment report (IPCC 2007; Table SPM 3) as one of the key technologies for reducing fugitive landfill emissions, i.e. methane that is not captured by collection systems and migrates to the atmosphere.

Previous studies conducted *in situ* have demonstrated the potential of biocovers to reduce methane emissions from landfills (e.g. Barlaz 2004; Cabral et al. 2009; Chanton and Liptay 2000; Humer and Lechner 1999; 2001; Jugnia et al. 2008; Stern et al. 2007; Whalen et al. 1990). In these studies, the substrate materials used, as well as methane loadings

applied, varied significantly. As part of the framework of a multidisciplinary study to assess the potential of passive methane oxidation biocovers (PMOB) to oxidize fugitive CH₄ under field conditions, three PMOBs were constructed within the existing final cover at the St-Nicéphore landfill, Quebec, Canada. Jugnia et al. (2008) analyzed the results obtained from PMOB-1, in which a mixture of sand and compost was used as substrate, and concluded that the mixture proved to be satisfactory in sustaining and promoting growth of methanotrophic bacteria. However, the methane loading into PMOB-1 was not controlled; as a consequence, it was not possible to determine the upper limit of methane loading that the biocover system would be able to oxidize.

The present study presents and analyses the results obtained during the 2008 monitoring campaign for PMOB-2 (the second system of the above-mentioned multidisciplinary study). For this particular PMOB, the same sand-compost mixture was used as substrate. However, the PMOB was lined with a geomembrane and biogas was fed through a gas distribution system, thereby allowing for control of the CH₄ loading. According to the pattern of results, the maximum capacity of the system does not seem to have been attained.

Materials and methods

Biocover setup

PMOB-2 (Fig. 1) measures 2.75m (W) × 9.75m (L) and follows a 3.5% slope. It was constructed within the existing final cover of the St-Nicéphore landfill, in an area where the waste mass is approximately 5-years old. PMOB-2 included a 0.80-m thick layer of substrate underlain by a 0.10-m thick gas distribution layer (GDL) consisting of 6.4-mm net gravel and 0.3 m of 12.7 mm net gravel. The substrate layer consists of a mixture of 5 volumes of

mature compost (before sieving through 12 mm industrial sieve) and 1 volume of coarse sand ($D_{10} = 0.07\text{mm}$; $D_{85} = 0.8\text{mm}$; $C_u = 4.3$). More details on the compost and the mixture can be found in Jugnia et al. (2008). The substrate layer was placed in four 0.2-m layers and compacted with a vibrating plate to obtain layers with an average dry unit weight of 8.4 kN m^{-3} and total porosity (n) equal to 0.63. The specific gravity (G_s) of the sand-compost mixture is equal to 2.24.

PMOB-2 was lined using a 1-mm thick HDPE geomembrane (GM), which was protected from tearing by a geotextile sheet. As a consequence, the biocover section was hydraulically isolated from the rest of the existing landfill cover, permitting the gas flow boundary conditions to be controlled, while maintaining the same atmospheric boundary conditions as the rest of the existing landfill cover. A drainage system was installed at the lowest point to evacuate infiltrating waters. The experimental plot was thermally shielded from the outside environment by 0.15-m thick polystyrene along its sides. The goal was to prevent lateral migration of moisture due to thermal gradients.

Instrumentation and Initial Conditions

Temperature probes (HOBO U12, from Onset) and water content capacitance sensors (ECH2O EC-5, from Decagon Devices) were installed at four depths throughout the cover at each of the four monitoring points (Fig. 2). The temperature and water content probes were connected to data loggers. At the same points, aluminum tubes equipped with a septum at the top end were inserted in the soil at 6 different depths (7 in some cases), in order to determine gas profiles (Fig. 3). Meteorological data, including precipitation, atmospheric pressure and wind speed were recorded continuously by a weather station installed near the experimental plot.

In the beginning of the study period (May to November 2008), the experimental plot was totally covered with thick vegetation, which frequently had to be removed manually in order to access the gas probes.

Gas Loading

The experimental plot was fed with biogas extracted from a well installed in the waste mass (Fig. 2). It was decided to use natural biogas from the site to maintain actual *in situ* conditions, i.e. the actual biogas mixture. We also wanted to keep intact the natural ratio of the stable isotopes ^{13}C and ^{12}C and content of odor-causing components of the biogas (for further investigations on odor control). The biogas loading (or fluxes) fed into the system was controlled by means of a valve. In the beginning of the study period, the CH_4 loading was measured using a rotameter, with readings being made once or twice a week. Subsequently, the CH_4 loading was monitored using a mass flow meter (Sage model SID-050-DC-24-DIG-GAS) connected to a data acquisition system. Samples for evaluation the CH_4 and CO_2 concentrations were taken from the well and end-cap of feed system.

Over the study period, the CH_4 load applied ranged from $0.3 \text{ g CH}_4 \text{ m}^{-2} \text{ h}^{-1}$ to $27 \text{ g CH}_4 \text{ m}^{-2} \text{ h}^{-1}$. The lower CH_4 loading value adopted corresponds to what is considered *ad hoc* as a residual loading that one would expect to find at the base of a landfill cover decades after closure of the site ($0.5 \text{ l CH}_4 \text{ m}^{-2} \text{ h}^{-1}$; or $0.3 \text{ g CH}_4 \text{ m}^{-2} \text{ h}^{-1}$; Stegmann et al. (2007)).

As indicated in Table 1, the study was subdivided into four periods corresponding to the four different CH_4 loadings. In the 1st period, the CH_4 loading was steadily increased from 0.3 to $8 \text{ g CH}_4 \cdot \text{m}^{-2} \cdot \text{h}^{-1}$, which was the maximum that could be monitored by the rotameter installed. During the 2nd period, the $8 \text{ g CH}_4 \cdot \text{m}^{-2} \cdot \text{h}^{-1}$ loading rate was maintained for several months to verify the stability of the system. Thereafter, all through the 3rd period, a mass flow

meter allowing monitoring of higher loadings was installed and the loading was again steadily increased up to 16 g CH₄. m⁻².h⁻¹. During the 4th period, the goal was to steadily increase the loading again so as to double the latter value. However, the experimental site had to be closed for the winter in early November, when the loading was equal to 27 g CH₄ m⁻².h⁻¹. As will be shown, the PMOB was capable of oxidizing all of the methane under this flux rate. This means that an upper limit on the amount of methane oxidation possible with this PMOB was not inferred in this study.

Gas Concentration Profiles and Emission Measurements

Emission measurements included CH₄ surface fluxes and surface scans of CH₄ concentration. CH₄ surface fluxes were measured following the static chamber method, at five permanent different points on the experimental plot (Fig. 2). The perimeter of the Plexiglas[®] chamber was sealed with a bentonite paste to prevent dilution of the gas inside the chamber by atmospheric air. CH₄ concentrations within the chamber were monitored every 10 seconds over a 6 minute interval using a portable FID (TVA 1000B, Thermo Scientific) equipped with a data acquisition system. Fluxes (*f*) were calculated according to the equation:

$$f = \frac{V}{A} \frac{\Delta C}{\Delta t} \quad (1)$$

where *f* is the CH₄ flux (mg/m²/s); *V* is the chamber volume (m³); *A* is the internal surface area (m²); and $\Delta C/\Delta t$ represents the slope of the plot relating change in gas concentration to time (mg/m³/s). The test was considered acceptable when the determination coefficient was equal or superior to 80% ($R^2 \geq 0.8$), as recommended by the U.S. Environmental Protection Agency (2003) for control of gas emissions from landfills.

CH₄ surface concentration scans were made using the FID, following a pre-defined path inside the PMOB. Gas samples were obtained continuously every 5 seconds, with a probe maintained at a distance of approximately 1 cm above the surface. The data was then used to define iso-concentration curves by kriging.

CH₄, CO₂ and O₂ concentration profiles within the PMOB were established from gas samples collected from the aluminum gas probes and analyzed *in situ* using a portable gas meter (Columbus Instruments Inc.) equipped with infrared sensors able to detect CO₂ and CH₄ on a scale from 0 – 100 vol% and an electrochemical sensor calibrated to detect O₂ from 0-21 vol%. The N₂ concentrations were obtained from the difference between 100% and the sum of the concentrations of CO₂, CH₄ and O₂. At various dates, the actual concentrations of N₂ were determined using a gas chromatograph (Micro GC 3000 A, Agilent).

Methanotroph Count

A previous study performed with samples from PMOB-1 (Cabral et al. 2007) showed that on average, the number of methanotrophic bacteria decreased quite abruptly within the first 0.4 m of depth from the surface. Based on these results, it was decided in the beginning of the 2008 monitoring season not to sample PMOB-2 for methanotroph count below the depth of 0.4 m. Substrate samples were collected monthly at three different points of the PMOB. The samples were taken from the uppermost part of the substrate (0-0.1, 0.1-0.2, 0.2-0.3 and 0.3-0.4 m) using PVC coring tubes (internal diameter = 0.5 m). Equal volumes of samples from the same depths were mixed to form composite samples that were used for methanotroph counts.

The cores were kept at 4°C and methanotroph counts were performed following the most probable number (MPN) method within 24 hours. For this, soil slurries from the

composite samples were serially diluted in 96-well plates (microtiter) containing ammonium mineral salts medium. Fresh soil (5 g) was suspended in 45 ml of a mineral salts medium (Heyer 2002) and shaken for 1 hour. Two hundred μl of this suspension was placed in the first well of a 96-well microtiter plate which consists of 12 wells. Serial 10-fold dilutions were performed from the second well to the 11th well with transfers of 20 μl to the next wells, containing 180 μl of mineral medium, using a multichannel pipettor. The 12th well contained only mineral medium and was used as the control. Thereafter, the plates were incubated for 4 weeks at 25°C in gastight jars containing 18% CH_4 in air. After incubation, all plates were read using a microplate reader and the MPN calculated from the dilution factor and dry weight of the soil.

Results and Discussion

Methane Removal Efficiency

Fig. 4 shows the evolution with time of the oxidation efficiency, CH_4 loading and emission rate. It can be observed that the CH_4 loading increased steadily up to mid-July, when it was left constant for a while. The associated outflow remained always close to nil or below detectable limits, which results in almost 100% efficiencies. By mid-September, the CH_4 loading was increased rapidly and a relatively higher outflow was measured on Oct. 3. But the system responded very well and the calculated efficiencies increased back again after Oct. 3 to values very close to 100%.

In Fig. 5, the relationship between CH_4 loading and absolute CH_4 removal rates can be observed. The CH_4 removal rate is equal to the difference between methane loading and surface emission. Fig. 5 shows that the absolute CH_4 removal rate increased with the

increasing loading; and the high determination coefficient obtained ($R^2 = 0.99$) indicates that the variability in the mass of CH_4 being supplied to the gas distribution layer below the biocover was the sole contributor to the variability in the CH_4 removal rate. Consequently, it is possible to anticipate that the maximum possible removal rate for this particular biocover was not reached at the end of this study.

Typical scans of methane concentrations at the surface of PMOB-2 are shown in Fig. 6. Due to the high oxidation efficiency of the biocover (Fig. 4), recorded concentrations of CH_4 at the surface remained quite low throughout the study; and were below the maximum concentration allowed by the Quebec landfill regulations, i.e. 500 ppm. The observed low concentrations of CH_4 along the PMOB-2's perimeter, as well as variability in the locations of registered peaks, can be interpreted as an indication that the seal along the interface between the substrate and the geomembrane was good enough to prevent gas leaks. As a consequence, the surface point measurements can be considered representative of the entire surface of the PMOB.

Gas profiles representative of the four periods into which the study was subdivided, are shown in Fig. 7. The profiles relative to the 1st period (Fig. 7a) showed that CH_4 was practically absent throughout the profile. The only plausible explanation for this observation is that CH_4 was being completely oxidized; even as deep as the interface between the substrate and the gas distribution layer (0.82 m). Indeed, N_2 concentrations at 0.82 m were similar to those found in atmospheric air, which means that the air supply – thus O_2 – was not a limiting factor. Nonetheless, the low O_2 concentrations deep down can be related to the fact that O_2 was immediately consumed as it became available.

The concentration of CH_4 at the bottom of PMOB-2 (0.82 m) gradually increased with the increase in the CH_4 inflow applied. Concurrently at this depth, there was a decrease in N_2

concentration, which means that the O_2 supply necessary for biotic degradation of CH_4 also decreased. The CH_4 concentrations normally found in landfill biogas (in the vicinity of 55%; Bogner et al. 1995; Börjesson et al. 2001) were observed at the bottom of PMOB-2, only when the CH_4 loading reached $27 \text{ g } CH_4 \text{ m}^{-2} \text{ h}^{-1}$ (Fig. 7d).

The steep declines in CH_4 concentrations shown in Fig. 7 b, c, d indicate that the oxidation front would be located quite deep in the PMOB (near a depth of 0.60 m) throughout the last three periods of CH_4 loadings. Humer and Lechner (2001) found, in a field test where sewage sludge and MSW compost were used as substrate, that the maximum CH_4 oxidation zone extended from 0.4 and 0.9 m deep.

It can be hypothesized that the presence of N_2 deep down for an extended period of time was enough of a proof that the incoming biogas was being diluted. However, it can be observed in Fig. 7 that the CO_2 concentration deep down was similar for periods 3 and 4. For this to happen CO_2 production must have taken place, otherwise the CO_2 would have been diluted to the same extent as the CH_4 .

The results presented above are quite different from those obtained for PMOB-1 (Rannaud et al. 2009), which differed from PMOB-2 only by the fact that it was not lined with a geomembrane. As a consequence, the gas distribution layer sits directly upon the waste mass and the biogas loading cannot be determined. Indeed, typical gas profiles for PMOB-1 show that CH_4 concentrations were usually high deep down (as compared to PMOB-2) and, with the exception of some short periods of draught, they remained high along the entire profile (Rannaud et al. 2009). Given the fact that the degrees of saturation of PMOB-1 and 2 were within the same range for all depths during the study period (see discussion below), one can conclude that very high upward biogas loadings in PMOB-1 (roughly estimated to be in

the order of $400 \text{ g CH}_4 \text{ m}^{-2} \text{ h}^{-1}$) must have prevented the downward migration of atmospheric O_2 , thereby precluding CH_4 oxidation.

In addition, in the beginning of the 2008 study period, the CH_4 loading applied to PMOB-2 was nil and thick vegetation (canary grass, peas, etc.) had developed on its surface. Given the fact that vegetation is known to increase rhizosphere-related microbial activity (e.g. Denier van der Gon 1996; Hilger and Humer 2003; Stralis-Pavese et al. 2004; Tanthachoon et al. 2008), it is highly possible that the establishment of vegetation had a positive impact on the performance of PMOB-2. However, this discussion is beyond the scope of the present paper, and no particular investigation was made concerning this matter.

Degree of Saturation within the Biocover

The evolutions of the degree of water saturation (S_r) within PMOB-2 and precipitation, during the study period, are presented Fig. 8. Only the average S_r values obtained for the two sub-layers, i.e. from 0.2 to 0.4 m and 0.6 to 0.8 m, are presented. The S_r values were calculated using volumetric water content data and the porosity ($n = 0.63$). As a crosscheck, these S_r values were compared to those obtained from a S_r vs suction plot (water retention curve of the material; data not shown), where suction measurements were obtained from tensiometers installed on the experimental plot (Fig. 3). A t-test analysis (results not shown) revealed no statistically significant difference between the two sets of data, i.e. between the calculated degrees of saturation and the degrees of saturation corresponding to measured suction values.

The degree of saturation of the substrate in PMOB-2 was sensitive to variations in the precipitation level. Indeed, peaks in precipitation levels associated with S_r values higher than 85% were observed after rainfall intensities equal to or greater than 20-25 mm/day; or

following continuous precipitation events. Otherwise, S_r values during the present study averaged $61.5 \pm 3.5\%$ at the topmost part of the PMOB (surface to 0.4 m deep) and $74.7 \pm 4.8\%$ within the bottom-most part (0.4 to 0.8 m). These average degrees of saturation values in the PMOB were then lower than 85%, which corresponds approximately to the degree of saturation value beyond which air becomes occluded in fine grained soils (Brooks and Corey 1966; Nagaraj et al. 2006). In addition, according to Nicholson et al. (1989), Yanful (1993), Aachib et al. (2004) and Cabral et al. (2004), gas fluxes become quite low when the value of S_r approaches 85%. Simulations performed by Rannaud et al. (2009) with S_r values in the vicinity of 85% resulted in very limited, to no penetration of O_2 , irrespective of the loadings applied. It can thus be inferred that high S_r would result in extremely low O_2 concentrations even at shallow depths, which would induce a significant decrease in microbial oxidation activity.

No correlation was found between S_r and the methane oxidation rate in the present study. It can thus be inferred that S_r values observed were within the optimal range for CH_4 oxidation to occur under the field conditions prevailing in PMOB-2.

It is relevant to note that the biocover substrate used in this experiment retains more moisture than most mineral soil covers; its volumetric water content varied between 32.0 and 63.0%, with this latter value occurring only very sporadically. Stern et al. (2007) concluded that biocover cells were more successful in oxidizing CH_4 when the retention times were long enough and desiccation was avoided as much as possible.

Temperature within the Biocover

Characteristic values of temperature obtained from the temperature probes placed in PMOB-2 indicated that temperatures in the top part of the PMOB (0.10 - 0.25 m) exceeded

the outside air temperature (Fig. 9). The combined effects of the warm landfill gas supplied and the heat generated by microbial activity are at the origin of this difference in temperature. However, when the air temperature started to drop, the bottom of the PMOB (0.45 – 0.70 m) became warmer than the top (Fig. 9), which is directly affected by outside weather conditions.

As shown in Table 2, there was only a small difference between average temperature values measured near the surface (20.1°C at 0.10 m) and deeper into the substrate (18.7 °C at 0.75 m). Gebert and Gröngroft (2006), who employed a much coarser substrate, obtained higher temperatures at the bottom of their biofilter experiment in the field. These authors suggested that the top layer is subjected to considerably higher, but also to considerably lower temperatures, presumably inducing higher and lower oxidation rates, than in the deeper layers. In the present study, the active oxidation zone seems to be located between 0.60 – 0.80 m, exactly within the region where a more constant temperature regime was observed (Fig. 9). The high temperatures at the top layers during the warmer months can be partly explained by aerobic activity and oxidation of organic matter. When the outside temperature started to drop, the bottom of the PMOB became warmer than the top (Fig. 9).

A statistical analysis performed using soil temperature and methane removal rate data led to an unusual negative correlation (-0.9; $p > 0.001$) between the two variables. Usually, methane removal rates (or for that matter, oxidation rates) increase with the increase in temperatures (e.g. Börjesson et al. 2004). In the present case, the statistical analysis does not reflect the physical phenomena, because the CH₄ loading was being increased precisely during the time of the year when the air temperature was starting to fall, causing a decrease in soil temperature, particularly near the surface (Fig. 9; the average temperature at the bottom of the PMOB was almost the same throughout the experiment). Nevertheless, despite

the decrease in soil temperature and the increase in CH₄ loading, almost 100% of the CH₄ was still being oxidized.

Gebert et al. (2003) claim that, during the winter, biofilters and biocovers may still be operated successfully, since, as the temperature in the soil drops, the methanotrophic population composition can adapt.

Methanotrophs and CH₄ Abatement

As shown in Fig. 10, the mean number of methanotrophs did not decrease considerably with depth, ranging from $7.82 \pm 4.45 \times 10^8$ CFU g_{dw}⁻¹ near the surface (0- 0.10 cm) to $1.73 \pm 1.14 \times 10^8$ CFU g_{dw}⁻¹ soil at the bottom (0.3 – 0.4 m). These densities are in the upper range of values reported in the literature and are comparable to those found by Gebert et al. (2003) (1.3×10^8 to 7.1×10^9 cells g_{dw}⁻¹) for a biofilter that was able to oxidize $80 \text{ g CH}_4 \text{ m}^{-3} \text{ h}^{-1}$, with absolute removal rates linearly correlated to the amount of methane entering the filter (such as illustrated in Fig. 5). Such uniform distribution profiles of methanotrophs suggest that the first 0.4 m of PMOB-2 offered optimal growth conditions. As mentioned previously (discussion relating to Fig. 7), air penetrated deep into the PMOB and, as a result, the molecular O₂ required by methanotrophs for CH₄ oxidation was not a limiting factor. Therefore, under the conditions that prevailed within PMOB-2 during this study, it is plausible to presume that the number of methanotrophs within the bottom layer (0.4 – 0.8 m; where samples were not collected from for microbial analyses) was as high as in the top layer. This would support the statement made previously that the potential removal rate obtained for PMOB-2 has possibly not been reached during this study.

Comparison of Biocover Performance to Methane Removal Rates Found for Other Biofilters and in Landfill Covers

Table 3 presents the CH₄ removal rates for several biofilter and biocover studies. The performance of the 0.8-m thick PMOB investigated in the present study, during which a removal rate of 27 g CH₄ m⁻² h⁻¹ (or 33.8 g CH₄ m⁻³ h⁻¹) was achieved under relatively low air temperatures (Fig. 9), is much higher than those found for the other selected landfill covers. However, the reported removal rates in Table 3 were limited to the low CH₄ loadings to which the covers were submitted in the field. It is therefore possible that higher removal rates might have been achieved had higher loadings be applied (unfortunately, monitoring had to be terminated because the site had to be closed for the winter). PMOB-2 was the only biocover in the list submitted to controlled CH₄ loading (biofilters are always submitted to controlled loadings).

The performance of PMOB-2 also compares rather well with the biofilters presented in Table 3, which were, with the exception of the one performed by Gebert and Gröngroft (2006), tested under the controlled conditions prevailing in the laboratory. Furthermore, if the compost added to the mixture used as substrate for PMOB-2 had not been sieved (as was the case with the sewage and waste compost used in Humer and Lechner's (2000) biocover study), a higher air-filled porosity would have been obtained, and the substrate would have been even more aerated, further facilitating CH₄ oxidation.

Final comments and conclusions

An experimental biocover constructed on top of the existing final cover at the St-Nicéphore landfill was tested under increasing CH₄ loadings. The results show that practically all the CH₄ provided to the system was oxidized, with a maximum removal rate of 27 g m⁻² h⁻¹ attained at the end of the study period, when the experimental site had to be closed for the winter. At this moment, a sharp decrease in temperature near the surface of the substrate

was being registered, while the air temperature varied widely during the day (reaching 0°C during the night), and the temperature deeper into the PMOB decreased, but not as steeply as near the surface.

The results also show that the absolute removal rates were linearly correlated to CH₄ loading, that the methanotrophs were present in great numbers near the surface, and that the oxidation zone was established between 0.6-0.8 m. In this bottom-most zone, we did not perform counts of methanotrophs, but we anticipate that they were present in great numbers given the conditions prevailing at these depths. All this leads to the conclusion that the maximum potential of PMOB-2 to oxidize CH₄ has not been reached. This is of great importance when considering that the oxidation rate attained far exceeds what has been considered as the residual loading one would expect to find at the base of a landfill cover decades after closure, i.e. 0.3 g CH₄ m⁻² h⁻¹.

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List of Tables

Table 1 – CH₄ loadings into PMOB-2

Table 2 - Minimum (Min), maximum (Max) and average temperature values within the BOPM-2

Table 3 – Methane removal rates by biofilters and landfill (bio)covers

List of Figures

Fig. 1. Cross Section of BOMP-2

Fig. 2. Position of the instrumentation profiles and surface flux measurement

Fig. 3. Instrumentation installed in PMOB-2

Fig. 4. Oxidation efficiency, methane loading and emissions as determined from surface emissions

Fig. 5. Methane removal rates as a function of CH₄ loading

Fig. 6. Representative surface scans of CH₄ concentrations for each loading period

Fig. 7. Representative gas profiles for each CH₄ loading period

Fig. 8. Evolution of (a) precipitation and degree of saturation in the PMOB-2 in 2008

Fig. 9. Evolution temperature within the PMOB-2 in 2008 (top and bottom sub-layers)

Fig. 10. Distribution with depth of the mean values of methanotroph counts within PMOB-2 for the study period

Table 1 – CH₄ loadings applied to PMOB-2 for each period

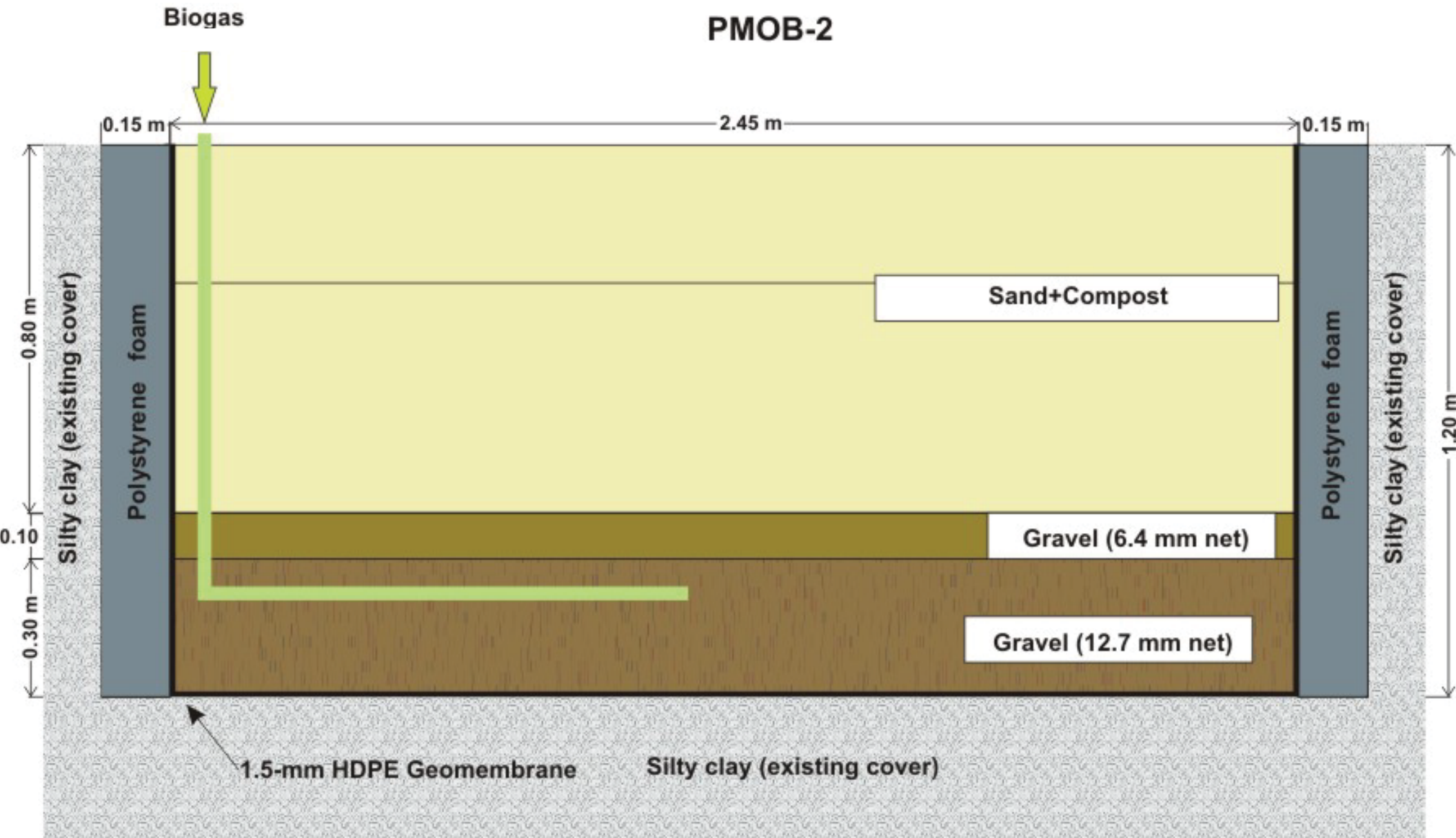
Time (in 2008)	Load (gCH ₄ m ⁻² h ⁻¹)
May to Mid-July	~ 0.3 to 8
Mid-July to September	~ 8
September to October	~ 16
October to November	~ 27

Table 2 - Minimum (Min), maximum (Max) and average temperature values within BOPM-2

Depth from Surface (m)	Average, °C	MAX, °C	MIN, °C
0 (air)	16.2	26	0
10	20.1	28.8	5.2
25	19.9	25.3	8.5
45	19.4	23.7	9.1
70	18.6	22.3	8.3

Table 3 – Methane removal rates by biofilters and landfill (bio)covers

Reference	Substrate material	System	Temp. (°C)	CH ₄ removal rate (g m ⁻³ h ⁻¹ for biofilters) (g m ⁻² h ⁻¹ for biocovers)
Whalen et al. (1990)	Loamy landfill cover	Biocover	25	2
Jones and Nedwell (1993)	Humic landfill cover	Biocover	22	0.3
Kightley et al. (1995)	Sandy landfill cover	Biocover	20	5–7
Humer and Lechner (2000)	Sewage sludge and waste composts	Biocover	18	1–16
Stern et al. (2007)	Pre-composted yard or garden waste placed	Biocover	25.7 ± 8.8	2
Sly et al. (1993)	Glass tubes	Biolfilter	-	21
Figueroa (1996)	Compost	Biolfilter	43	50
Streese and Stegmann (2003)	Compost	Biolfilter	30	65
Wilshusen et al. (2004)	Compost	Biolfilter	Room temperature	4.1
Melse and Var der Werf (2005)	Compost - perlite	Biolfilter	12	11–15
Gebert and Gröngroft (2006)	Expanded clay	Biolfilter	3 – 24 (avg 12 °C° in the soil)	Max 80
This study (PMOB-2)	Sand/Compost	Biocover	See Fig. 9	27 (maximum not reached; test discontinued when site had to be closed for the winter)



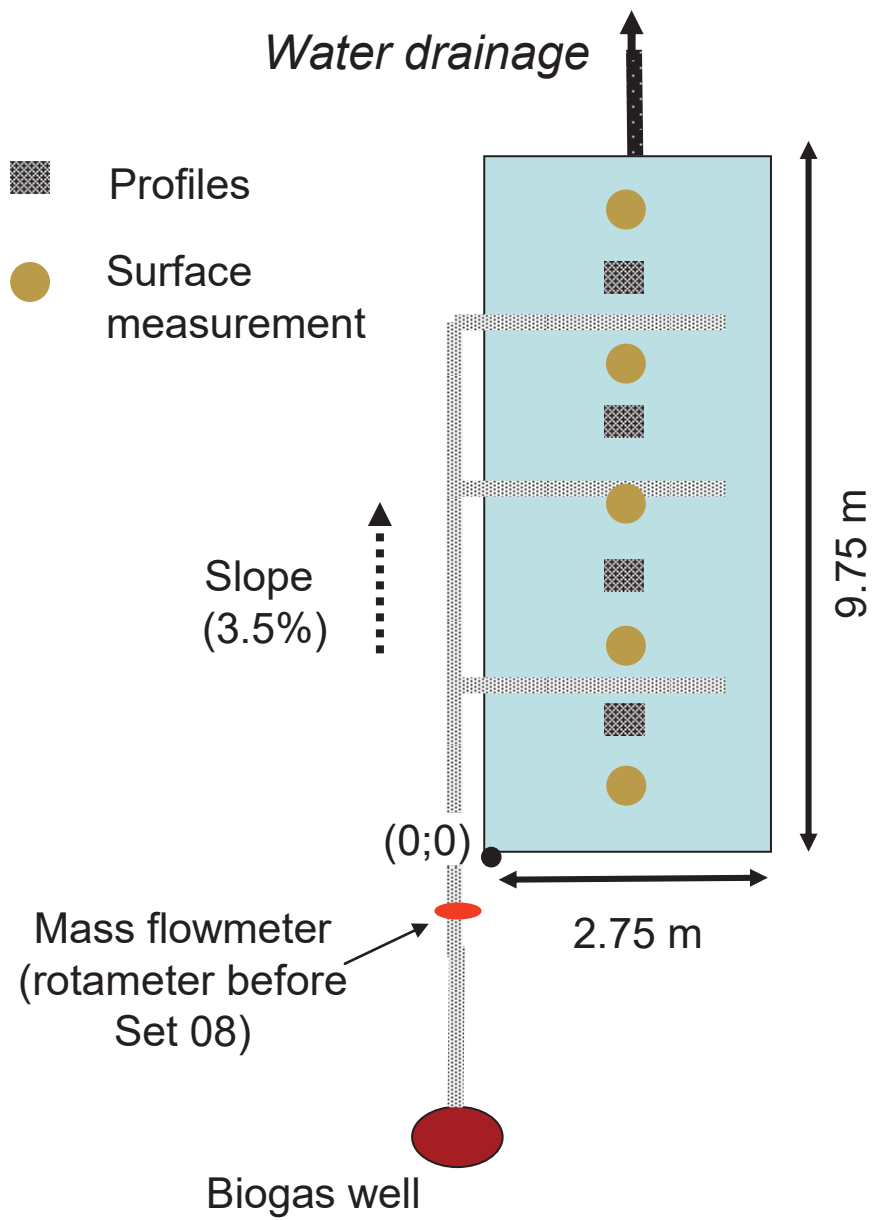


Fig. 2. Position of the instrumentation profiles and surface flux measurement

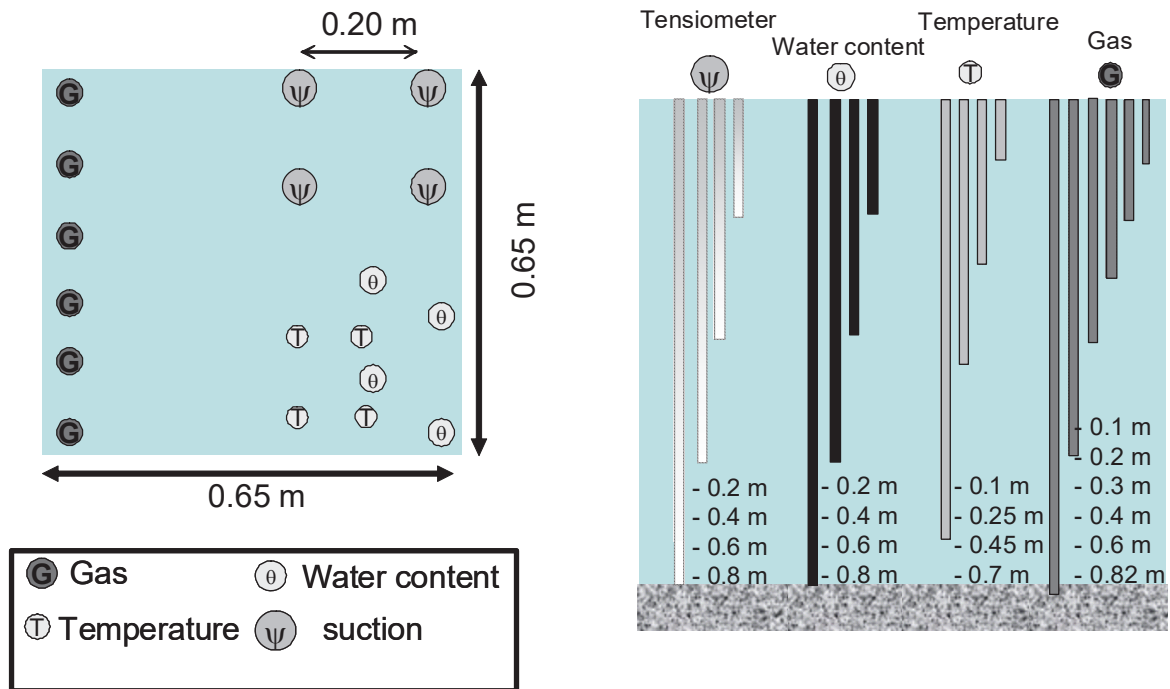


Fig. 3. Instrumentation installed in PMOB-2

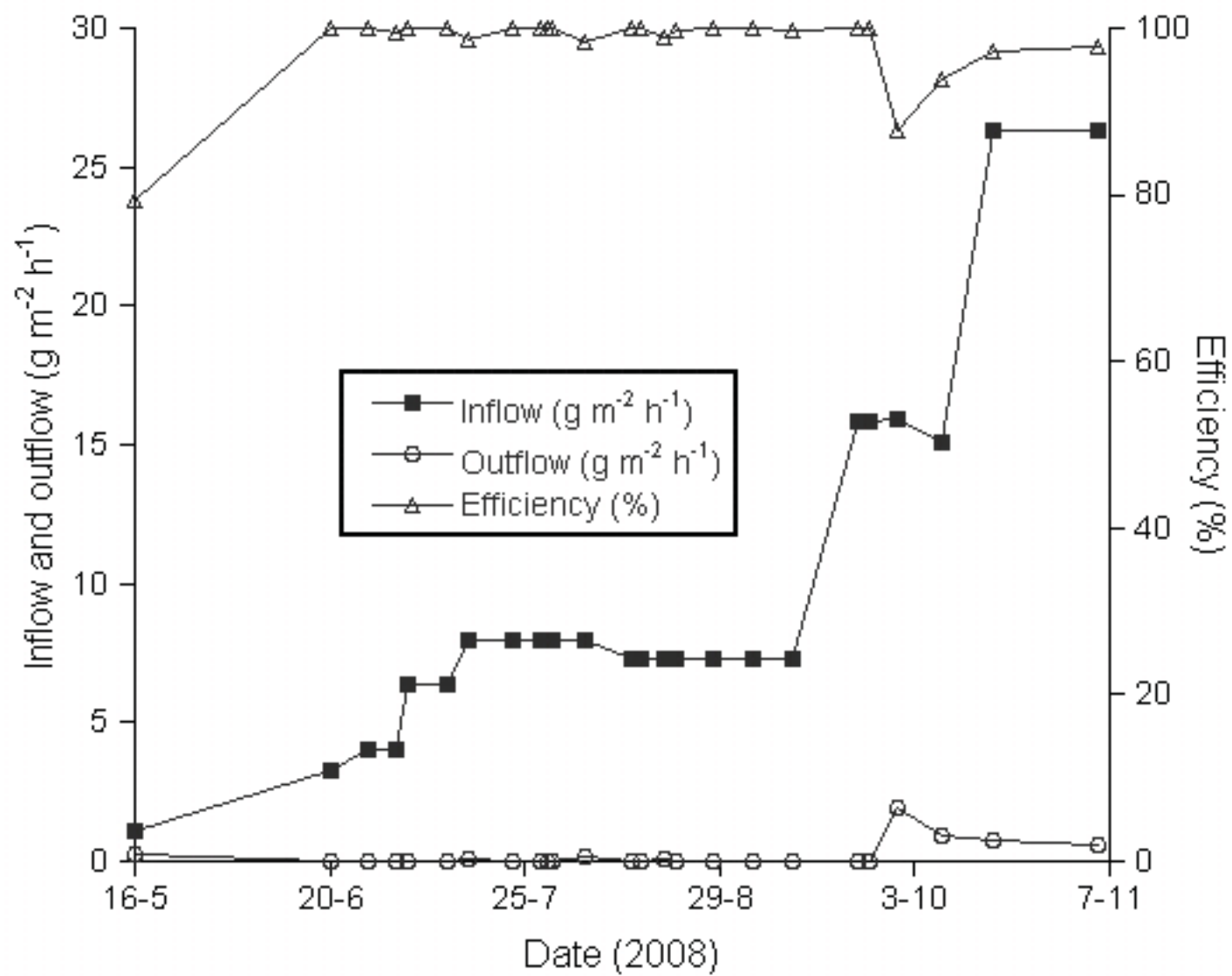


Fig. 4. Oxidation efficiency, methane loading and emissions as determined from surface emissions

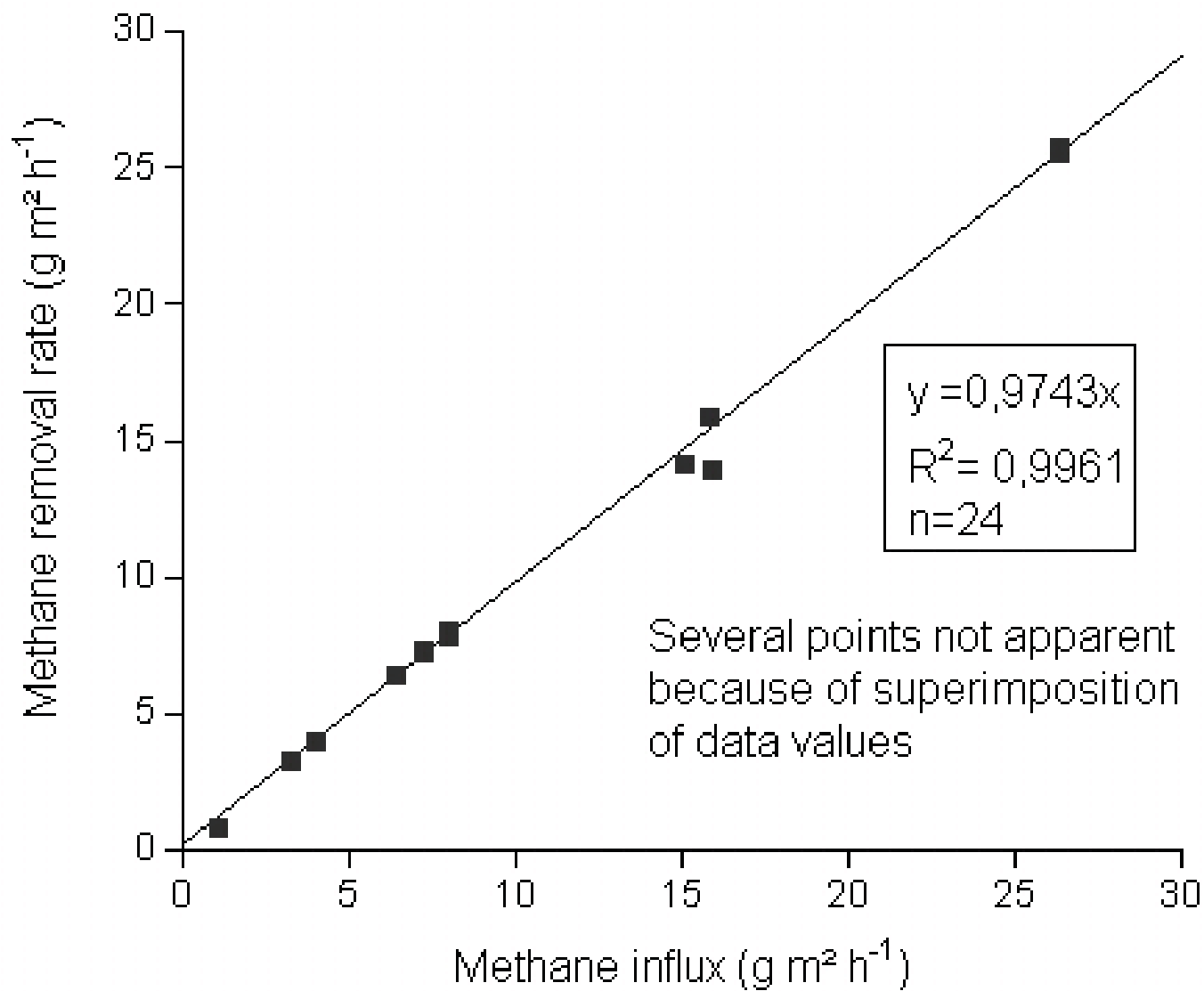


Fig. 5. Methane removal rates as a function of CH₄ loading

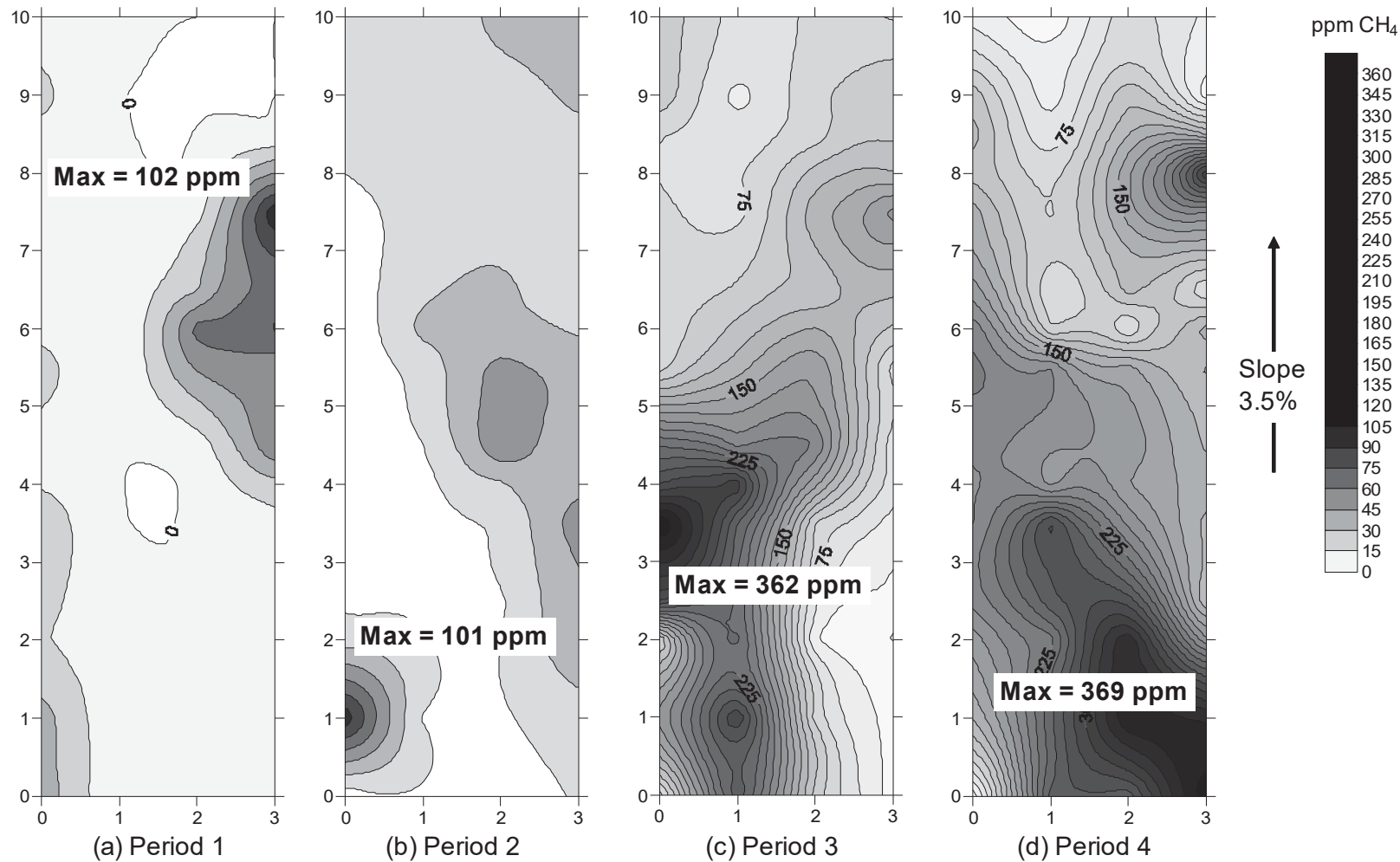


Fig. 6. Representative surface scans of CH₄ concentrations for each loading period

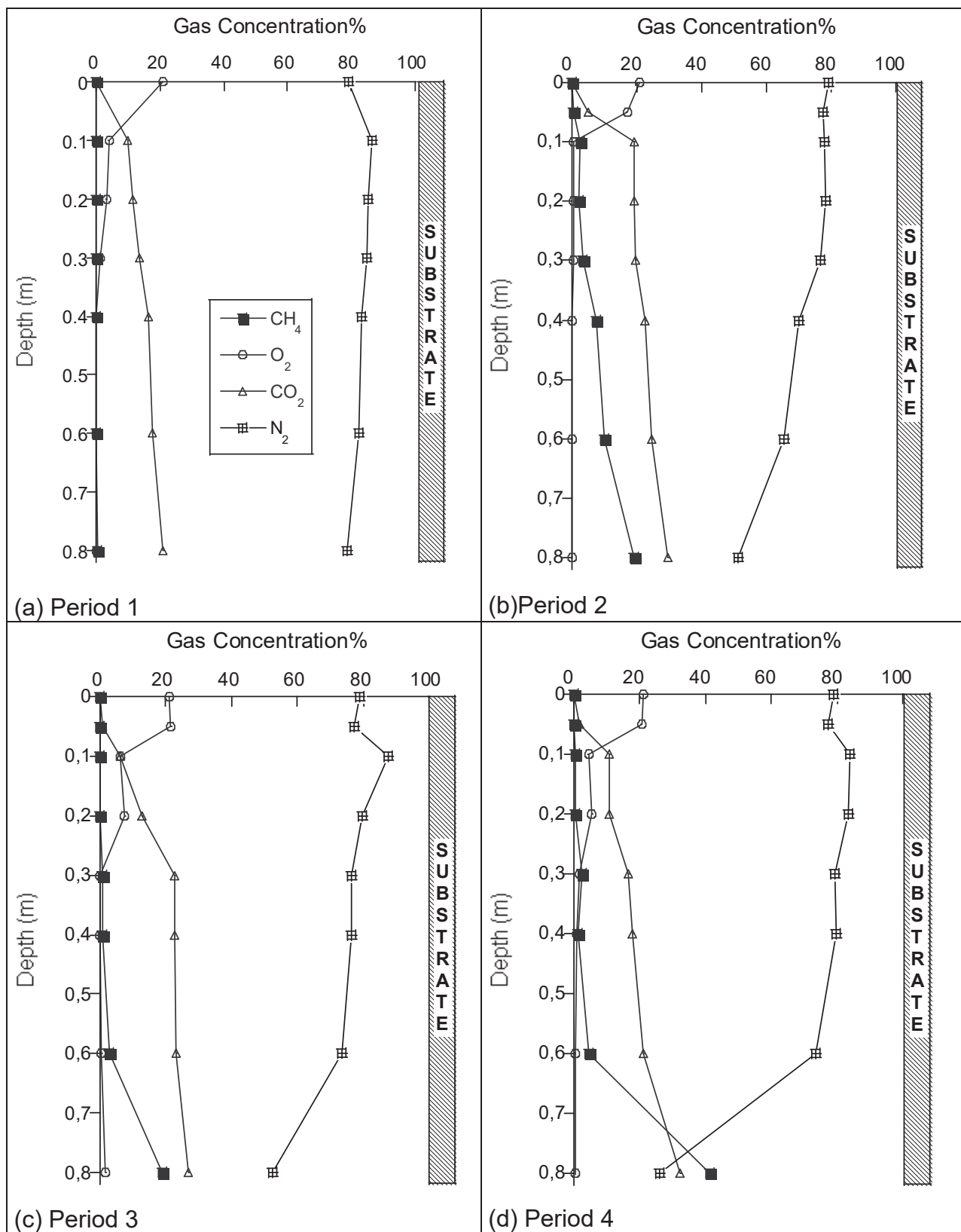


Fig. 7. Representative gas profiles for each CH₄ loading period

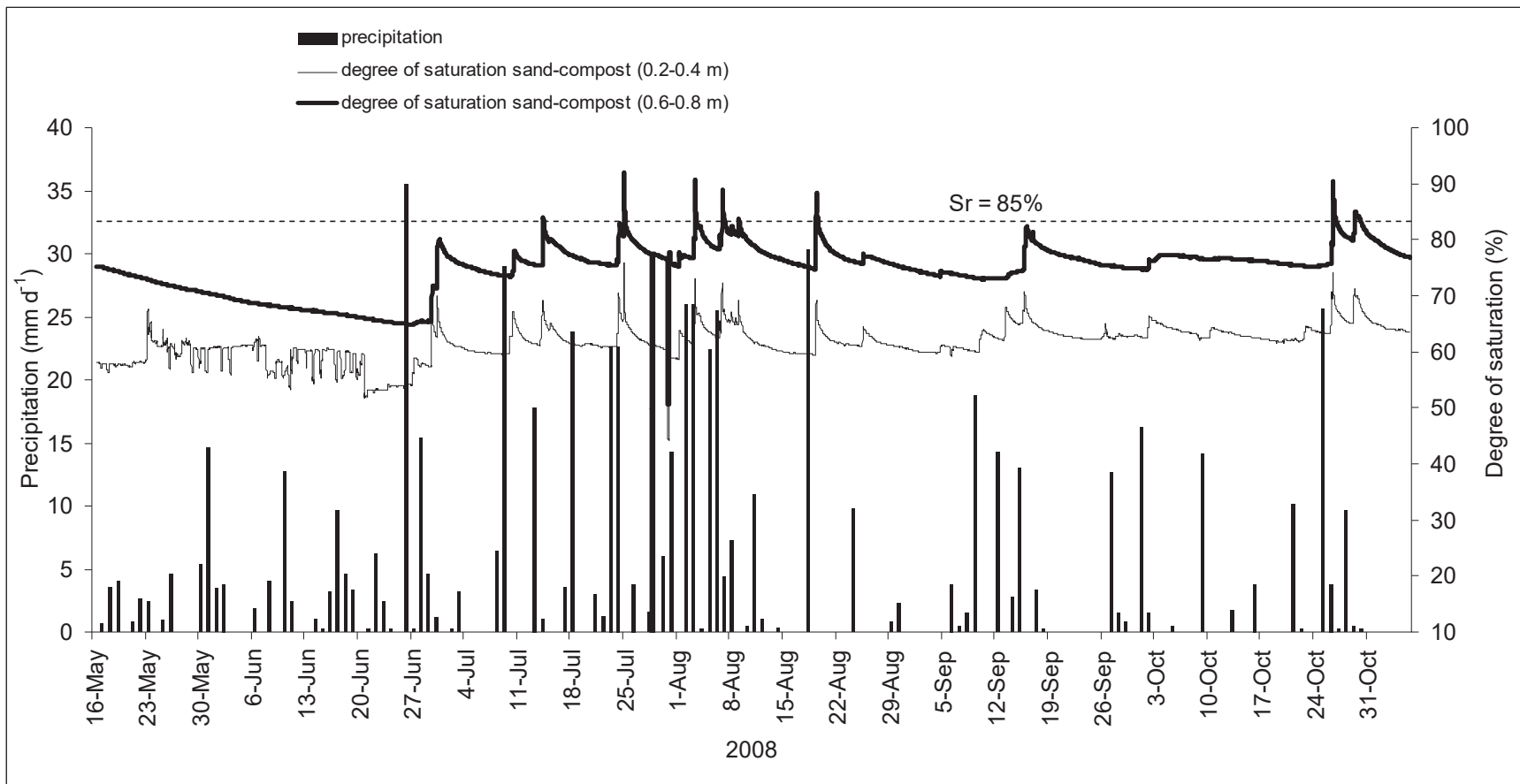


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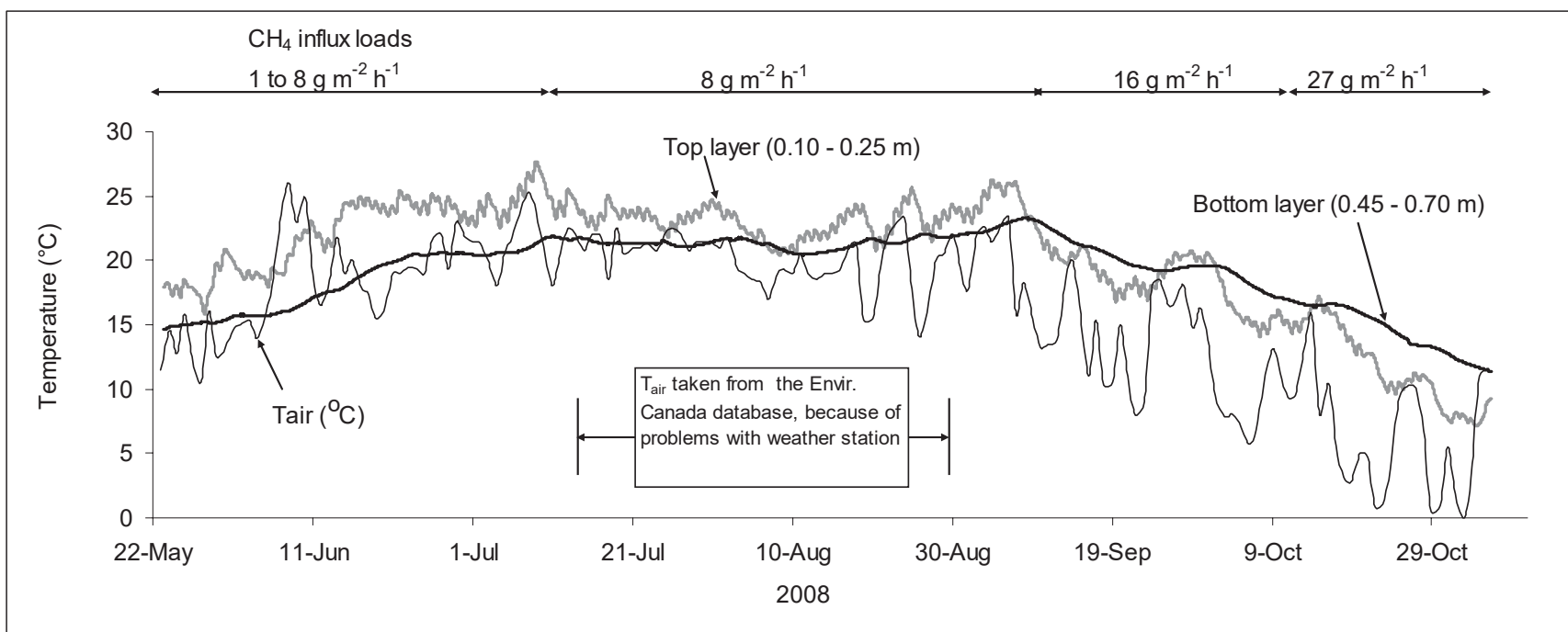


Fig. 9. Evolution of temperature within the PMOB-2 in 2008 (top and bottom sub-layers)

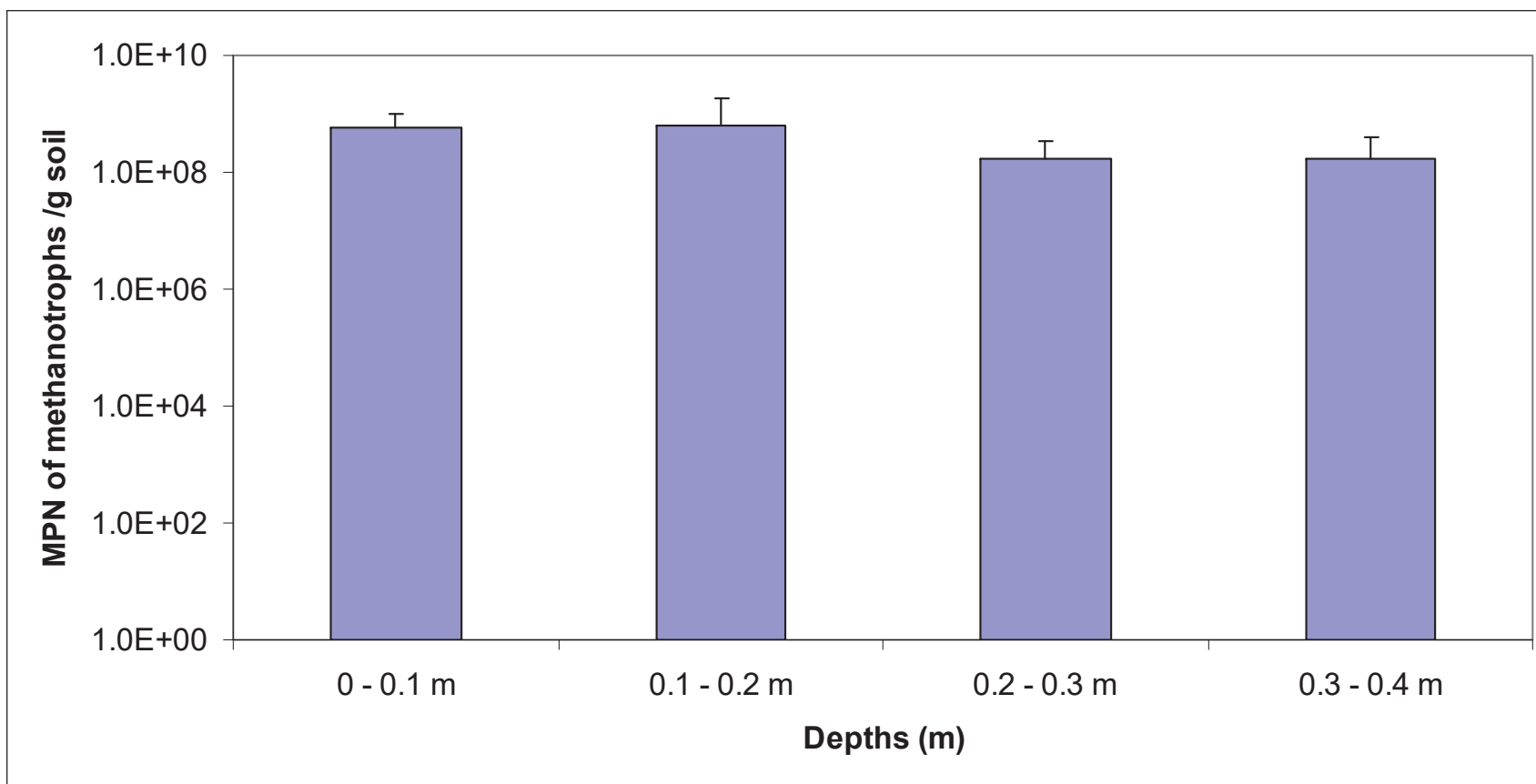


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